

Prevalence of *Escherichia Coli* Pathotypes in Stools of HIV-Positive Adults Attending a HAART Clinic Ileife, Nigeria

¹Eniola, K. I. T., ²Awoniyi, L. M., ³Torimiro, N, ¹Ajayi O. O. and ¹Adeyemo-Eleyode V. O.

¹Environmental and Public Health Research Laboratory, College of Agriculture and Natural Sciences, Joseph Ayo Babalola University, Ikeji-Arakeji, Osun State, Nigeria.

²Virology Research Clinic, Obafemi Awolowo University Teaching Hospital Complex, Ile Ife.

³Department of Microbiology, Obafemi Awolowo University, Ile Ife

DOI: <u>https://doi.org/10.51244/IJRSI.2024.1109044</u>

Received: 26 August 2024; Revised: 03 September 2024; Accepted: 07 September 2024; Published: 05 October 2024

ABSTRACT

Morbidity and mortality among HIV/AIDS patients are often exacerbated by opportunistic infections (OIs). *Escherichia coli* has emerged as a significant agent of OIs in HIV-positive patients. The use of highly active antiretroviral therapies (HAART) has improved health outcomes, including a reduction in OIs like diarrhoea. This study aimed to identify and characterize the pathotypes of *E. coli* present in the stools of HIV-positive adults attending the HAART Clinic in IIe-Ife, Nigeria. *E. coli* was isolated and pathotyped based on the presence of specific virulence genes: *eaeA*, *bfpA*, *stx1*, *stx2*, *eltB*, *estA*, *ipaH*, *pCVD*, and *EcoRI-PstI*. Out of 271 stool samples examined, *E. coli* was detected in 27 (9.96%). Among the 72 *E. coli* strains isolated, 21 were identified as diarrheagenic *E. coli* (DEC): 11 (52.38%) were Enteropathogenic *E. coli* (EPEC), 7 (33.33%) were Shiga toxin-producing *E. coli* (STEC), and 3 (14.29%) were Enterohemorrhagic *E. coli* (EHEC) pathotypes. Notably, five stool samples contained more than one DEC pathotype. The study found a low prevalence of DEC, with three major pathotypes prevalent among the HIV-positive adults sampled. Identifying specific *E. coli* pathotypes in HIV/AIDS patients is crucial for understanding potential complications and associated risks, which can guide clinicians in selecting appropriate treatments and managing bacterial co-infections more effectively.

Keywords: Diarrheagenic *Escherichia coli* (DEC), Opportunistic infections (OIs), HAART (Highly Active Antiretroviral Therapy).

INTRODUCTION

Opportunistic infections (OI) are a significant cause of morbidity and mortality in people with HIV infection due to their weakened immune systems (Wingfield and Wilkins, 2010). HIV destroys the immune system and renders patients susceptible to opportunistic infections (Elfstrand and Floren, 2010). HIV Infection causes a decline in the levels of CD4 T cells and affects critical cells (macrophages and dendritic cells) in the body system (Kumar, 2018; Cunningham *et al.*, 2020; Ponnan *et al.*, 2021; Chang *et al.*, 2022). There is a progressive loss of protective antibodies, which often causes the patients to become a microbial zoo (Geoff et al., 2022; Arora and Arora, 2009).

Kibwengo *et al.* (2022) opined that opportunistic infections (OIs) are the first clinical manifestations that alert clinicians to the occurrence of acquired immunodeficiency syndrome (AIDS). In their study, they reported that delay in antiretroviral treatment (ART) initiation after positive test results, poor drug adherence and moderate malnutrition are major risk factors that affect the occurrence of OI. The establishment of opportunistic infection in all circumstances depends on quality of treatment and exposure to infectious agents (*Mycobacterium tuberculosis, Candida albicans, Escherichia coli, Cryptosporidium parvum, Toxoplasma gondi* and so on) (Kibwengo *et al.*, 2022; Shrestah *et al.*, 2022; Tan *et al.*, 2012)



The microbial agents associated with OI include bacteria, fungi, viruses, and protozoa, which can cause opportunistic infections, such as pulmonary, oropharyngeal, gastrointestinal, dermatological, neurological, ophthalmic and even multi-system infections. Among these, gastrointestinal infections represent a serious public health risk (Wingfield and Wilkins, 2010). Diarrhoea is a common problem in HIV-infected individuals in many developing countries (Carcamo *et al.*, 2005; Sun *et al.*, 2022). Dysentery and bacterial gastroenteritis are among opportunistic infections (OIs) often associated with HIV patients (Carcamo *et al.*, 2005; Okeke, 2009 and DeWitt *et al.*, 2019). E. coli is widely associated with diarrhoea. Braz *et al.* (2020) categorised *E. coli* into diarrheagenic *E. coli* (DEC), non-pathogenic *E. coli*, and extraintestinal pathogenic *E. coli*. They further subdivided the extraintestinal pathogenic *E. coli* (NMEC). DEC has been consistently implicated as a cause of diarrhoea in adults infected with HIV14- (Oluma and Richard, 2008; Zhu *et al.*, 2019 and Verma *et al.*, 2022).

One of the goals of Highly Active Antiretroviral Therapy (HAART), a combined therapy, is to improve immune function (Eggleton and Nagalli, 2022). The recovery of immune status with HAART has markedly improved the long-term outcome for patients with adequately treated HIV infection because it inhibits viral replication and greatly reduces the chances of treatment failure or development of multidrug resistance (Eggleton and Nagalli, 2022).

Although HAART has resolved diarrhoea cases caused by opportunistic pathogens previously considered untreatable; MacArthur and Duport (2012) indicated that most diarrhoea cases in HIV-infected patients on HAART are not due to opportunistic infection but are due to protease inhibitors. Treating diarrhoea in HIV-infected patients will be difficult if the specific cause is not known. Therefore, this study was designed to investigate the pathotypes of *E. coli* in adult HIV-positive patients on HARRT.

MATERIALS AND METHODS

Study population and sample collection

The study involved 271 consented HIV positive adult patients aged between 18 and 60 years who are on HAART. Ethical clearance (number: IRB/IEC/0004553 NATIONAL: NHREC/27/02/2009a) was obtained for the study from the Ethics and Research Committee of the Obafemi Awolowo University Teaching Hospital Complex, Ile-Ife.

Stool samples were collected from participants during their clinic days into sterile universal bottles. The stool samples were analysed within 30 minutes at the Virology Research Clinic (VRC) of the Obafemi Awolowo University Teaching Hospital Complex (OAUTHC) between August and October 2021.

Isolation and identification of Escherichia coli

E. coli was isolated by streaking the stool sample onto sterile Eosin Methylene Blue (EMB) agar plates (Oxoid Ltd, Basingstoke Hampshire, England) using the quadrant streak plate method (Poyil *et al.*, 2022) and incubated at 37°C for 24 hours. Typical *E. coli* colonies with greenish metallic sheen on EMB agar were sub-cultured and further characterised by standard biochemical tests. Genomic DNA of each isolate was extracted as described by Dashti *et al.* (2009) and their identity confirmed by PCR using specific primer (New England *BioLabs*) (Bej *et al.*, 1991; Aranda *et al.*, 2020)

Screening for diarrheagenic Escherichia coli

All the *E. coli* isolates were screened for virulence genes characteristic of Enterotoxigenic *E. coli* (ETEC), Enteroinvasive *E. coli* (EIEC), Enteropathogenic *E. coli* (EPEC), Enteroaggregative *E. coli* (EAEC) and Enterohaemorrhagic *E. coli* (EHEC) (Aranda *et al.*, 2004; Abongo et al., 2018; Momba *et al.*, 2008). The target genes were *eaeA* (structural gene for intimin of EHEC and EPEC), *bfpA* (structural gene for the bundle-forming pilus of EPEC), *stx1* and/or *stx2* (shiga toxins 1 and 2 of EHEC and STEC), *eltB* and/or *estA* (enterotoxins LT and ST of ETEC), *ipaH* (invasion-associated locus of the invasion plasmid found in EIEC and *Shigella*) and pCVD (the nucleotide sequence of the *EcoRI-PstI* DNA fragment of pCVD432 of EAEC). *E. coli* strains



E2348/69, O42, H10407, EDL 933 and E137 served as positive controls for EPEC, EAEC, ETEC, EHEC and EIEC respectively.

Statistical analysis

Data were analysed using WinPepi version 11.65. Descriptive statistics (Percentage, Frequency and Mean) of data was presented. Chi-square test was used to test for the association between the groups and a P < 0.05 was considered statistically significant.

RESULTS

E. coli was isolated from 27 (9.96 %) of the 271 stool samples analysed, some of the samples yielded more than one *E. coli* isolate and a total of 72 *E. coli* strains were confirmed. Among the 72 *E. coli* strains, 21 were found to be DEC strains while the other 51 were non-DEC strains. Three pathotypes of DEC: EPEC, STEC and EHEC were detected. The most prevalent was EPEC (11; 52.38 %), two of its sub-pathotype: Typical EPEC (7: 33.33 %) and Atypical EPEC (4: 19.05 %) were detected (Table 1). More than one DEC pathotype was recovered from 5 of the stool samples. All the Atypical EPEC strains carried the *eae* genes, while six of the Typical EPEC carried the *bfp* genes, and one of the Typical EPEC carried the *bfp* and *eae* genes. Two of the STEC strains carried *stx1* genes, three of them carried *stx2* genes, one carried *stx1* and *bfp*, while another one carried *stx2* and *bfp*. Two of the EHEC carried *bfp*, *eae*. and *stx2* genes while one other carried *eae*, and *stx2* genes.

Sample Code	Number of DEC in samples	Pathotypes	Virulence Genes
10	2	Atypical EPEC	Eae
		Typical EPEC	Bfp
12	2	STEC	stx2
		Typical EPEC	Bfp
37	1	Typical EPEC	Bfp
68	1	Atypical EPEC	Eae
70	1	Atypical EPEC	Eae
88	2	EHEC	stx2, eae, bfp
		Typical EPEC	eae,bfp
99	1	STEC	stx2, bfp
102	2	STEC	stx1, bfp
		Typical EPEC	Bfp
103	1	STEC	stx2
133	1	STEC	stx1
169	1	Typical EPEC	Bfp
184	1	STEC	stx1

Table 1 Pathotypes of Diarrheagenic E. coli (DEC) in Stool of HIV Positive Adults



186	1	STEC	stx2
207	2	Atypical EPEC	Eae
		EHEC	stx2, eae
268	1	EHEC	stx2, eae,bfp
271	1	Typical EPEC	Bfp

Keys: EPEC: Enteropathogenic E. coli EHEC - Enterohemorrhagic E. coli

STEC: Shiga toxin-producing E. coli

DISCUSSION

The frequency of *E. coli* in the stools of the HIV patients (9.00%) in this study is higher than the 1.9% reported by Reuben and Gyar (2015). However, it was lower than the 12.16%, 42% and 18% reported by Abongo *et al.* (2008), Garcia *et al.* (2010), and Ngalani, *et al.* (2019) respectively. Although most strains of *E. coli* are commonly facultative bacteria in the lower intestine of humans and warm-blooded animals, they may act as opportunistic pathogens (Braz *et al.*, 2020). *E. coli* is gradually becoming a serious public health concern as an opportunistic infection associated with people living with HIV in developing countries (Okeke, 2009; DeWitt *et al.*, 2019 and Chabala *et al.*, 2020), the low frequency of its detection in the subjects suggests that it is either not yet a widespread problem or is being managed successfully.

DEC constituted a smaller proportion (29.17%) of *E. coli* isolated in this study compared with 39% obtained by Olaru etc al., 2021. The three pathotypes of DEC detected in this study have been associated with HIV positive patients by Alizade *et al.* (2017). EPEC which is the most frequent pathotype (15.28%), in the diarrheagenic adult HIV patients involved in this study, was also the most prevalent (30.77%) in the study by Medina *et al.* (2010), This appears to buttress the view that EPEC strains are becoming significant enteropathogens in immunocompromised adults (Alizade *et al.*, 2017). STEC which is the second most prevalent (9.72%) DEC pathotype detected in this study has been reported by Alizade *et al.* (2019). The low frequency of EHEC pathotype detection in this study is similar to the result obtained by Kebede *et al.* (2020); which detected only two EHEC strains in 102 stool samples from HIV-positive adults. In the study by Llorente *et al.* (2023); Okeke and Nataro (2001) reported EAEC pathotype in the stools of HIV infected adults, however, the pathotype was not isolated in this study.

CONCLUSION

The study provides valuable data on the prevalence and types of *E. coli* pathotypes among HIV/AIDS patients around Ile-Ife, Nigeria, which is useful for targeted interventions by public health authorities to reduce bacterial infections among this vulnerable population. Information on specific *E. coli* pathotypes in HIV/AIDS patients, which the study provides is useful in understanding the potential complications and risks associated with these infections and can guide clinicians in selecting appropriate treatments and managing bacterial co-infections more effectively. This study reported a low prevalence of DEC in HIV-positive adults with a preponderance of EPEC and STEC pathotypes. Most of the isolates were resistant to a range of antibiotics. The study also highlights the need for continuous monitoring of bacterial infections in HIV-positive individuals, particularly in regions with high HIV prevalence.

REFERENCES

1. Abongo, B. O., Okomo, G. and Ouma, C. 2018. Community Acceptability of Voluntary Medical Male Circumcision (VMMC) as a Strategy in the Fight against the Spread of HIV and AIDS among Residents of Homa-Bay County Kenya. Texila International Journal, 6(3): 9.



- 2. Alizade, H., Sharifi, H., Naderi, Z., Ghanbarpour, R., Bamorovat, M. and Aflatoonian, M. R. 2017. High frequency of diarrheagenic Eschericia coli in HIV-Infected and Patients with thalassemia in Kerman, Iran. Journal of International Association of Providers of AIDS Care, 16(4): 353-358.
- Aranda, K.R., Fagundes-Neto, U., and Scaletsky, I.C. A. (2020). Evaluation of Multiplex PCRs for Diagnosis of Infection with Diarrheagenic Escherichia coli and Shigella spp. Journal of Clinical Microbiology, 42 (12): 5849-5853.
- 4. Arora, D. R., and Arora, B. (2009). AIDS-associated parasitic diarrhea. Indian Journal of Medical Microbiology, 27(3), 185-190. https://doi.org/10.4103/0255-0857.53199
- 5. Bej, A. K., McCarty, S. C., and Atlas, R. M. (1991). Detection of coliform bacteria and Escherichia coli by multiplex polymerase chain reaction: comparison with defined substrate and plating methods for water quality monitoring. Applied and environmental microbiology, 57(8), 2429-2432.
- 6. Braz, V. S., Melchior, K. and Moreira, C. G. 2020. Escherichia coli as a multifaceted pathogenic and versatile bacterium. Frontiers in Cellular and Infection Microbiology, 10: 548492.
- Cárcamo, C., Hooton, T., Wener, M.H., Weiss, N.S., Gilman, R., Arevalo, J., Carrasco, J., Seas, C., Caballero, M., and Holmes, K.K. (2005). Aetiologies and manifestations of persistent diarrhea in adults with HIV-1 infection: a case-control study in Lima, Peru. Journal of Infectious Diseases. 191:11–19. https://doi.org/10.1086/426508
- 8. Chabala, F., Madubasi, M., Mutengo, M. M., Banda, N., Yamba, K., and Kaonga, P. (2020). Escherichia coli antimicrobial susceptibility reduction amongst HIV-infected individuals at the university teaching hospital, Lusaka, Zambia. International Journal of Environmental Research and Public Health, 17(10), 3355.
- Chang, J. H., Chuang, H. C., Hsiao, G., Hou, T. Y., Wang, C. C., Huang, S. C., and Lee, Y. L. (2022). Acteoside exerts immunomodulatory effects on dendritic cells via aryl hydrocarbon receptor activation and ameliorates Th2-mediated allergic asthma by inducing Foxp3+ regulatory Tcells. International Immunopharmacology, 106, 108603. http://doi.org/10.1016/j.intimp.2022.108603
- Cunningham, S., and Hackstein, H. (2020). Recent Advances in Good Manufacturing Practice-Grade Generation of Dendritic Cells. Transfusion Medicine and Hemotherapy, 47(6), 454-463. https://doi.org/10.1159/000512451
- 11. Dashti, A. A., Jadaon, M. M., Abdulsamad, A. M., and Dashti, H. M. (2009). Heat treatment of bacteria: a simple method of DNA extraction for molecular techniques. Kuwait Med J, 41(2), 117-122.
- 12. DeWitt, J. C., Blossom, S. J., and Schaider, L. A. (2019). Exposure to per-fluoroalkyl and polyfluoroalkyl substances leads to immunotoxicity: epidemiological and toxicological evidence. Journal of exposure science & environmental epidemiology, 29(2), 148-156.
- 13. Eggleton, J. S. and Nagalli S. (2022) Highly Active Antiretroviral Therapy (HAART) Stat Pearls https://www.ncbi.nlm.nih.gov/books/NBK554533/
- 14. Elfstrand, L. and Florén C. (2010) Management of chronic diarrhea in HIV-infected patients: current treatment options, challenges, and future directions. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3218691/pdf/hiv-2-219.pdf
- 15. Garcia, C., Chincha, O., Leon, M., Iglesias, D., Barletta, F., Mercado, E., and Ochoa, T. (2010). High frequency of diarrheagenic Escherichia coli in human immunodeficiency virus (HIV) patients with and without diarrhea in Lima, Peru. The American journal of tropical medicine and hygiene, 82(6), 1118.
- 16. Groff, D., Carlos, N. A., Chen, R., Hanson, J. A., Liang, S., Armstrong, S., Li, X., Zhou, S., Steiner, A., Hallam, T. J. and Yin, G. 2022. Development of an E. coli strain for cell-free ADC manufacturing. Biotechnology and Bioengineering, 119(1): 162-175.
- 17. Kebede, A., Tessema, F., Bekele, G., Kura, Z. and Merga, H. 2020. Epidemiology of survival pattern and its predictors among HIV positive patients on highly active antiretroviral therapy in Southern Ethiopia public health facilities: a retrospective cohort study. AIDS Research and Therapy, 17(1): 49.
- Kibwengo, C. F., Kabalami, T. K. and Sungwa, E. 2022. Opportunistic Infections and Associated Factors among HIV Infected Patients on Antiretroviral Treatment at Bombo Hospital in Tanga Region, Tanzania. African Journal of Health Sciences, 35(3): 252-262.
- 19. Kumar, B. V., Connors, T. J., and Farber, D. L. (2018). Human T cell development, localisation, and function throughout life. Immunity, 48(2), 202-213. https://doi.org/10.1016/j.immuni.2018.01.007
- 20. Llorente, M. T., Escudero, R., Ramiro, R., Ramacha, M. A. Martinez-Ruiz, R., Gala-Sandez, F., de Frutos, M., Elia, M., Onnubia, I. and Sanchez, S. 2023. Enteroaggregative Escherichia coli as etiological



agent of endemic diarrhea in Spain: A prospective multicenter molecular characterization of isolates. Frontiers in Microbiology, 14: 1120285.

- MacArthur, R. D. and DuPont, H. L. (2012). Etiology and Pharmacologic Management of Noninfectious Diarrhea in HIV-Infected Individuals in the Highly Active Antiretroviral Therapy Era. Clinical Infectious Diseases. 55(6), 860–867, https://doi.org/10.1093/cid/cis544
- 22. Medina, A. M., Rivera, F. P., Romero, L. M., Kolevic, L. A., Catilo, M. E., Verne, E., Harnandez, R., Mayor, Y. E., Barletta, F., Mercado, E. and Ochoa, T. J. 2010. Diarrheagenic Escherichia coli in Human Immunodeficiency Virus (HIV) pediatric patients in Lima, Peru. The American Journal of Tropical Medicine and Hygiene, 83: 158-163.
- 23. Momba, M. N. B., Abong'o, B. O. and Mwambakana, J. N. 2008. Prevalence of enterohaemorrhagic Escherichia coli O157:H7 in drinking water and its predicted impact on diarrhoeic HIV/AIDS patients in the Amathole District, Eastern Cape Province, South Africa. Water SA, 34(3): 365-372.
- 24. Ngalani, O. J. T., Mbaveng, A. T., Marbou, W. J. T., Ngai, R. Y. and Kuete, V. (2019) Antibiotic resistance of enteric bacteria in HIV-infected patients at the Banka Ad-Lucen hospital, West Region of Cameroon. Canadian Journal of Infectious Diseases and Medical Microbiology, 2: 9381836.
- 25. Okeke, I. N. (2009). Diarrheagenic Escherichia coli in sub-Saharan Africa: status, uncertainties and necessities. The journal of infection in developing countries, 3(11), 817-842.
- 26. Okeke, I. N. and Nataro, J. P. 2001. Enteroaggregative E. coli. The Lancet Infectious Diseases, 1: 304-313.
- 27. Oluma, Y.B., and Richard, L.G. (2008). Aetiology and Management of Diarrhea in HIV-infected Patents and Impact on Antiretroviral Therapy. Global HIV/AIDS Medicine. Pg: 737 745.
- 28. Ponnan, S. M., Vidyavijayan, K. K., Thiruvengadam, K., Hilda J, N., Mathayan, M., Murugavel, K. G., and Hanna, L. E. (2021). Role of Circulating T Follicular Helper Cells and Stem-Like Memory CD4+ T Cells in the Pathogenesis of HIV-2 Infection and Disease Progression. Frontiers in immunology, 12, 1179. https://doi.org/10.3389/fimmu.2021.666388
- 29. Poyil, M. M., Karuppiah, P., Raja, S. S., and Sasikumar, P. (2022). Isolation, Extraction, and Characterization of Verotoxin-producing Escherichia coli O157: H7 from Diarrheal Stool Samples. Sudan Journal of Medical Sciences, 17(1), 116-127.
- Reuben, C. R., and Gyar, S. D. (2015). Isolation and antibiogram of shiga toxin-producing Escherichia coli O157: H7 from diarrhoeic HIV/AIDS patients in Lafia, Central Nigeria. Int. Res. J. Microbiol, 6, 20-26. http://dx.doi.org/10.14303/irjm.2015.135
- 31. Shrestah, R., Maviglia, F., Altice, F. L., DiDomizio, E., Khati, A., Mistler, C., Azwa, I., Kamarulzaman, A. Halim, M. A. A., and Wickersham, J. A. 2022. Mobile Health Technology Use and the Acceptability of an mHealth Platform for HIV Prevention Among Men Who Have Sex with Men in Malaysia: Crosssectional Respondent-Driven Sampling Survey. Journal of Medical Internet Research, 24(7): e36917.
- 32. Sun, C. J., Shato, T., Steinbaugh, A., Pradeep, S., Rivet Amico, K., and Horvath, K. (2022). Virtual voices: examining social support exchanged through participant-generated and unmoderated content in a mobile intervention to improve HIV antiretroviral therapy adherence among GBMSM. AIDS care, 1-9. <u>https://doi.org/10.1080/09540121.2022.2038364</u>.
- 33. Tan, J. Y., Huedo-Medina, T. B., Warren, M. R., Carey, M. P. and Johnson, B. T. 2012. A meta-analysis of the efficacy of HIV/AIDS prevention interventions in Asia, 1995–2009. Social Science and Medicine, 75(4): 676-687.
- 34. Wingfield, T. and Wilkins, E. 2010. Opportunistic infection in HIV disease. British Journal of Nursing, 19(10): 621-627.
- 35. Verma, S., Venkatesh, V., Kumar, R., Kashyap, S., Kumar, M., Maurya, A. K., and Singh, M. (2019). Etiological agents of diarrhea in hospitalised pediatric patients with special emphasis on diarrheagenic Escherichia coli in North India. Journal of Laboratory Physicians, 11(01), 068-074. https://doi.org/10.1177%2F17562848221092593
- 36. Zhu, Z., Zhao, R., and Hu, Y. (2019). Symptom clusters in people living with HIV: a systematic review. Journal of Pain and Symptom Management, 58(1), 115-133. https://doi.org/10.1016/j.jpainsymman.2019.03.018.